Please replace the paragraph at Page 18, lines 13-21 with the following rewritten paragraph:

Moreover, methods for randomly introducing mutations are not particularly limited, and Mutagenic PCR as described below can preferably be used in this invention. The Mutagenic PCR can be carried out according to methods known in the art. (C.W. Dieffenback, ed. PCR PRIMER, A Laboratory Manual (Cold Spring Harbor Laboratory Press) (1995) pp. 583-588.) Concretely, the following conditions were employed in the examples.

A4

Please replace the paragraph at Page 18, line 22 through Page 19, line 8 with the following rewritten paragraph:

About 50 ng of Plasmid BlueBFP (201) was added to 10 × mutagenic PCR buffer (70 mM $MgCl_2$, 500 mM KCl, and 100 mM Tris-HCl, pH 8.3 at 25 °C; 0.1% (w/v) gelatin) 10 µl, 10 × dNTP (2 mM dGTP, 2 mM dATP, 10 mM dCTP, and 10 mM dTTP) 10 µl, 10 mpol/µl primer (23mer M13Universal primer and M13Reverse primer) 3µ1, and H_2O 62µl, and mixed. Subsequently, 10 µl of 5mM $MnCl_2$ was added and mixed, and 1µl of Taq Polymerase (Takara) was added to conduct PCR (PC-700 available from ASTEC Inc. was used). The PCR was conducted in three tubes under the following conditions: 25 cycles at 94 °C for 1 min, 30 cycles at 45 °C for 1 min, and 35 cycles at 72 °C for 1 min, respectively.

A5

Please replace Table 4 on Page 23, lines 5-20 with the following revised table:

TABLE 4

GFP						
101	none					
103			Phe64Leu			
104				Val163Ala	Ser175Gly	
105			Phe64Leu,	Val163Ala,	Ser175Gly	
BFP (as for BFP, the two mutations, Tyr66His (Y66H) and Tyr145Phe (Y145F), have been introduced into the sequence for GFP which serves as a basis)						
201	Y66H,	Y145F:				
202	Y66H,	Y145F:	Phe64Leu,			Leu236Arg
203	Y66H,	Y145F:	Phe64Leu			
204	Y66H,	Y145F:		Val163Ala,	Ser175Gly	
205	Y66H,	Y145F:	Phe64Leu,	Val163Ala,	Ser175Gly,	Leu236Arg

Please replace the paragraph at Page 25, line 19 through Page 26, line 19 with the following rewritten paragraph:

Unless otherwise so stated, CHO-K1 cells were grown in a F12+10% FBS medium in 5% CO_2 at 37 °C. The cells (1×10^5) were inoculated into a 6-cm dish, and on the following day, their transfection was conducted in two dishes as a pair by the calcium phosphate method. (C. Chen and H. Okayama Mol. Cell. Biol. 7: 2745-2752 (1987).) After transfection, the one dish was incubated at 37 °C and the other at 30 °C for 24 h. The transfected CHO cells were washed with 1 × PBS (-) three times, and they were dissolved in 1 ml of 10 mM Tris-HCl (pH 7.4)

containing 1% Triton X-100 and recovered in an Eppendorf tube. A supernatant (0.5 ml) from centrifugation at 3,000 rpm for 5 min was diluted 4-fold with the same buffer and fluorescence measurement was performed. Here, a pUcD2SRαMCS vector (empty vector) was transfected and used as a blank. A Hitachi F-2000 type fluorophotometer was used in the fluorescence measurement. In the measurement of GFPs, fluorescence was scanned between 460 nm and 600 nm at an excitation wavelength of 460 nm to measure the maximal value in the vicinity of the fluorescence wavelength of 510 nm. In the measurement of BFPs, fluorescence was scanned between 360 nm and 500 nm at an excitation wavelength of 360 nm to measure the maximal value in the vicinity of the fluorescence wavelength of 445 n.

A7 cncld.

Please replace the paragraph at Page 26, line 21 through Page 27, line 22 with the following rewritten paragraph:

The CHO cells were transfected with pUcD2SRαMCS (empty vector)(T. Tsukamoto et al. Nature. Genet. 11: 395-401 (1995)), phGFP(101)-Cl, phGFP(105)-Cl, phBFP(201)-CL, and phBFP(205)-Cl, respectively and grown at 37 °C and at 30 °C. Employing a sample prior to dilution as used in the fluorescence measurement previously described (8 µl), SDS-PAGE was performed on a 12% gel. With the use of a Horizonblot (ATTO Inc.), transfer was conducted onto a nitrocellulose membrane (Millipore Inc., HAHY394FO) under the conditions of 2 mA and 90 min per cm^2. After the membrane was taken out and washed with 1 × PBS, it was immersed in 1% skim milk/PBS and shaken at room temperature for 30 min. After the membrane was washed with 1 × PBS, it was immersed in 0.1% skim milk/PBS containing an anti-GFP antibody (Clonetech Inc.) that had been diluted 2,000-fold and shaken at 4 °C overnight. The membrane was washed with 1 × PBS for 5 min, and then with TPBS (0.05% Trion X-100/PBS) for 15 min three times. The membrane was immersed in 0.1% skim milk/PBS containing an anti-rabbit IgG antibody labeled with HRP (Amersham Inc.) that had been diluted 1,000-fold, and shaken at 4

A8 cn.t